

# Antioxidant and anti-inflammatory activities of Gallic acid in Japanese quails induced by oxidative stress

## Actividades antioxidantes y antiinflamatorias del ácido gálico en codornices japonesas inducidas por estrés oxidativo

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### ABSTRACT

Gallic acid is a phenolic compound found in many plant sources with strong antioxidant activity. In this study, the bioactivity of Gallic acid was investigated in Japanese quails induced by oxidative stress. The study was performed on four groups of 40-day-old male Japanese quail (*Coturnix japonica*). Oxidative stress was created for 1 week by adding 0.5% hydrogen peroxide. The study was terminated by administering 100 mg·kg<sup>-1</sup> body weight Gallic acid intraperitoneally. Total antioxidant and total oxidant level analyzes from liver tissue homogenates were performed using a ready-made commercial kit. TNF- $\alpha$  levels from blood samples taken for anti-inflammatory activity were investigated by ELISA method. There were no statistically significant results on live weight gain between the experimental groups and control group. However, Gallic acid in liver homogenates together with H<sub>2</sub>O<sub>2</sub> increased total antioxidant state (TAS) compared to H<sub>2</sub>O<sub>2</sub> application, while it decreased total oxidant state (TOS) in the same groups. Moreover, while the oxidative stress index increased in the H<sub>2</sub>O<sub>2</sub> group, it decreased significantly in both the Gallic acid and Gallic acid + H<sub>2</sub>O<sub>2</sub> groups. Gallic acid application also caused regression in blood TNF- $\alpha$  expression levels, which were increased by H<sub>2</sub>O<sub>2</sub>. In quails, Gallic acid showed antioxidant activity by increasing TAS levels and decreasing TOS levels, providing a significant decrease in oxidative stress index. It also provided anti-inflammatory activity by suppressing TNF- $\alpha$  levels. However, advanced molecular analyzes are needed to obtain more detailed information on the subject.

**Key words:** Gallic acid; growth performance; oxidative stress; inflammation; quails

### RESUMEN

El ácido gálico es un compuesto fenólico que se encuentra en muchas fuentes vegetales con fuerte actividad antioxidante. En este estudio, se investigó la bioactividad del ácido gálico en codornices japonesas inducidas por estrés oxidativo. El estudio se realizó en cuatro grupos de codornices japonesas macho (*Coturnix japonica*) de 40 días de edad. Se creó estrés oxidativo durante 1 semana mediante la adición de peróxido de hidrógeno al 0,5 %. El estudio finalizó con la administración de 100 mg·kg<sup>-1</sup> de peso corporal de ácido gálico por vía intraperitoneal. Los análisis del nivel de antioxidante total y oxidante total de homogeneizados de tejido hepático se realizaron utilizando un kit comercial ya preparado. Los niveles de TNF- $\alpha$  de muestras de sangre tomadas para determinar la actividad antiinflamatoria se investigaron mediante el método ELISA. No hubo resultados estadísticamente significativos sobre la ganancia de peso vivo entre los grupos experimentales y el grupo control. Sin embargo, el ácido gálico en homogeneizados de hígado junto con H<sub>2</sub>O<sub>2</sub> aumentó el estado antioxidante total (TAS) en comparación con la aplicación de H<sub>2</sub>O<sub>2</sub>, mientras que disminuyó el estado oxidante total (TOS) en los mismos grupos. Además, mientras que el índice de estrés oxidativo aumentó en el grupo H<sub>2</sub>O<sub>2</sub>, disminuyó significativamente, tanto en el grupo de ácido gálico como en el de ácido gálico + H<sub>2</sub>O<sub>2</sub>. La aplicación de ácido gálico también provocó una regresión en los niveles de expresión de TNF- $\alpha$  en sangre, que aumentaron con H<sub>2</sub>O<sub>2</sub>. En codornices, el ácido gálico mostró actividad antioxidante al aumentar los niveles de TAS y disminuir los niveles de TOS, proporcionando una disminución significativa en el índice de estrés oxidativo. También proporcionó actividad antiinflamatoria al suprimir los niveles de TNF- $\alpha$ . Sin embargo, se necesitan análisis moleculares avanzados para obtener información más detallada sobre el tema.

**Palabras clave:** Acido gálico; desempeño del crecimiento; estrés oxidativo; inflamación; codornices

## INTRODUCTION

Recently, especially after the limitation of the use of antibiotics, the use of alternative natural compounds, which are abundant in plants and fungi and have important biological activities, as natural feed additives in poultry farming has been increasing. Polyphenols constitute the most effective groups of phytochemicals added to animal feeds as an alternative to antibiotics [1, 2]. Polyphenols are defined as compounds containing at least one hydroxyl group attached to a phenol ring in their chemical structure. The most widely known of these compounds, which have a wide range of varieties, are Caffeic acid, Ferulic acid, p-Hydroxybenzoic acid, Protocatechic acid, Vanillic acid, Salicylic acid and Gallic acid [3, 4].

Gallic acid (3,4,5-Trihydroxybenzoic acid), an important member of the tannins group of these phenolic compounds, attracts the attention of researchers in scientific studies due to its strong antioxidant bioactivity [5], although Gallic acid, or gallate, is found in many plants [6] and in fungal species [7]. In addition to the antioxidant activity of this compound, which was discovered many years ago, anticancer [8], anti-HIV [9], antiulcerogenic [10], anti-inflammatory [11], antimicrobial [12] and antifungal [13] have also been reported in studies.

Oxidative stress basically develops when free radicals cannot be adequately removed by antioxidant defense mechanisms. Normally, there are antioxidant defense mechanisms in order to maintain the oxidant-antioxidant balance in living things. When this defense system, which includes enzymatic mechanisms such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and nonenzymatic mechanisms such as glutathione, vitamin C, vitamin E and plant polyphenolic compounds, is weakened, the production and accumulation of reactive oxygen species in cells and tissues increases [14]. Oxidative stress is one of the main reasons for the deterioration of animal health and reduced meat quality in poultry farming [15]. Phytochemicals found in plants, especially polyphenols, stand out as the most important supplement compounds in coping with oxidative stress thanks to their strong antioxidant activities [16]. Therefore, these compounds have the potential to be used as feed additives to reduce oxidative stress in poultry and increase productivity in animals [17]. This study was aimed to investigate some parameters of antioxidant and anti-inflammatory activities of Gallic acid in a hydrogen peroxide-induced oxidative stress model in quails (*Coturnix japonica*).

## MATERIALS AND METHODS

### Ethical approval

This study was approved by Hatay Mustafa Kemal University Animal Experiments Local Ethics Committee (Decision No: 2021/06-17).

### Experimental design

The study was performed on 40-day-old male Japanese quail. The quails were individually weighed on a precision scale (Ohaus NV622, USA), and their average body weight was recorded and distributed to the experimental groups (n=10).

In the study groups, the control group (C) was fed only commercial starter feed, group I was administered commercial feed starter + Gallic acid intraperitoneally (IP), group II was given commercial feed starter and hydrogen peroxide (HP) added to drinking water, finally group III was applied commercial feed starter + HP + Gallic acid IP. Oxidative stress was created for 1 week by adding 0.5 % hydrogen peroxide

(Tekkim, Turkey) to the drinking water of quails in groups II and III. Then, the study was terminated by administering Gallic acid (Sigma Aldrich, EU), dissolved in serum physiologic at a concentration of 100 mg·mL<sup>-1</sup> body weight intraperitoneally for 2 days to the experimental groups I and III every other day.

In the study, quails were fed with commercial growth feed (TABLE I). In the study, first body weight and final body weight, 10-day Body Weight change rate (%) and Average Daily Feed Intake Amount g were measured (n=10). Afterwards, tissue and blood samples were taken and oxidative stress parameters and biochemical-molecular analyzes were performed (n=5).

**TABLE I**  
Nutrient composition of commercial grower feed

Nutrient values	Amount (%)
Crude protein	20.0
Crude Fat	3.00
Crude fiber	3.00
Crude ash	4.80
Lysine	1.12
Methionine	0.51
Calcium	0.880
Total phosphorus	0.440
Sodium	0.140
Vitamin–Mineral premix*	0.250
Metabolizable energy kcal·kg <sup>-1</sup>	2859.3

\*1 kg of the premix provided: 15,000,000 IU of Vitamin A, 5,000,000 IU of Vitamin D3, 100,000 mg of Vitamin E, 3000 mg of Vitamin K3, 5000 mg of Vitamin B1, 8,000 mg of Vitamin B2, 60,000 mg of niacin, 15,000 mg of D-calcium pantothenate, 5000 mg of Vitamin B6, 20 mg of Vitamin B12, 200 mg of D-biotin, 2000 mg of Folic acid, 100,000 mg of Vitamin C, 0.02 mg of Cyanocobalamin, 74 mg of Mn (from MnO), 45 mg of Zn (from ZnO), 4 mg of Cu (from CuO), 12.5 mg of Fe (from FeSO<sub>4</sub>), 0.3 mg of I (from KI), 0.15 mg of Se (from NaSe)

### Determination of biochemical parameters

In the study, liver samples were taken from 5 quails from each group for biochemical parameters. Total antioxidant status (TAS) and total oxidant status (TOS) analyzes from tissue homogenates (Rel assay-TR) were performed using a ready-made commercial kit, following the methods of Kucukgul and Erdogan [18]. The Oxidative Stress Index (OSI) in the experimental groups was calculated by the ratio of these two values. In addition, Tumor necrosis factor alpha (TNF-α) levels from blood samples taken for anti-inflammatory activity were investigated by ELISA method with commercial kits (Kit Origin).

### Statistical analysis

Statistical analysis with the help of IBM SPSS 22 package program, the comparison of the groups in terms of the examined features was analyzed with One-way Anova. Differences were determined by Duncan's test.

## RESULTS AND DISCUSSIONS

Studies on performance and egg quality of Gallic acid and its derivatives in poultry (*Gallus gallus domesticus*), especially in layers, are quite limited. However, when reviewing the literature, it is seen that there are studies using grape (*Vitis viniferas*) seed extract gallic acid abundant in the seeds and skin of the grape. For example, Abdel-Wahab *et al.* [19] reported that grape seed polyphenols decreased total cholesterol lipid levels, blood sugar and liver enzyme activities, and increased glutathione peroxidase enzyme activity in Japanese quails, especially in 10 to 38-day periods. Again, Silici *et al.* [20] reported that the addition of 0.5, 1 and 1.5 % ground grape seed to quail compound feeds did not have a negative effect on yield, hatchability and egg quality of quails, and also increased feed efficiency. Moreover, Abu Hafsa and Ibrahim [21] reported that with the addition of 20 g·kg<sup>-1</sup> grape seed (GS) to the basal diet, the final live weight and live weight gain of quails increased, their feed conversion ratio increased, but feed intake was not affected. However, they stated that meat significantly increased carcass physical and chemical composition properties, carcass yield, and gizzard percentage. They reported that the addition of 40 g·kg<sup>-1</sup> GS significantly reduced the percentage of abdominal fat in poultry. In addition, in a study by Ao and Kim [22], it was reported that the addition of grape seed extract increased feed efficiency, increased antioxidant enzyme activities and immunity, and improved meat quality and body weight gain in Peking ducks (*Anatidae*).

In the present study, the live weight change rates of quails in the experimental groups (group I, II and III) were found to be lower (4.45 %, 4.25 % and 2.69 % respectively) than the control group (10.52 %). Also, according to the results obtained, feed conversion ratios between the experimental groups and the control group were not found statistically significant. In addition, when the experimental groups were compared among themselves, it was determined that the live weight gain rates changed in parallel with the feed intake amount.

Oxidative stress, known as the disruption of the oxidant-antioxidant balance between reactive oxygen species and the body's antioxidant defense systems, constitutes one of the most fundamental problems of modern poultry farming [23]. Increasing superoxide and hydroxyl radicals as a result of the disruption of this balance cause serious damage to lipid, deoxyribonucleic acid (DNA), protein and other cellular components, leading to disruption of cell integrity and tissue damage [24]. It is known that oxidative stress affects commercial poultry production, growth performance, productivity and reproductive performance of layer hens in poultry farming. Although there are various strategies for the control of oxidative stress in poultry, natural antioxidant compounds (polyphenols), which are non-toxic and do

not leave residues, have recently come into the focus of scientific studies [25]. The health benefits of plant phenolics are mainly due to their antioxidant and anti-inflammatory abilities [26]. Among such plant phenolics, Gallic acid (a trihydroxybenzoic acid) found in various foods and plants can be given as an example [27]. The curative effects of Gallic acid on many metabolic diseases including obesity have been reported [28, 29]. Moreover, several reviews have been published focusing on the therapeutic potential of Gallic acid. In 2013 Locatelli *et al.* [30] studied alkyl esters of Gallic acid as anticancer agents. Again, Choubey *et al.* [31] demonstrated the anticarcinogenic, antimicrobial, antimutagenic and antiangiogenic properties of Gallic acid and its esters.

When the liver total antioxidant capacity values were compared, TAS value in group I (0.9748 ± 0.15944) was found to be 5 % lower than the control group (1.0306 ± 0.12777), whereas in group II (0.6401 ± 0.09142) decreased by 37 % (P<0.5). However, this value increased 55 % in group III (0.9971 ± 0.13702) compared to group II (P<0.5) (FIG. 1). According to the results that was obtained in the present study, the TAS levels of Japanese quails decreased with the application of hydrogen peroxide, while the application of hydrogen peroxide together with Gallic acid prevented this decrease and increased the TAS value to the levels of the control group. In a study, Samuel *et al.* [32] showed that while Gallic acid decreased MDA, it increased plasma SOD and TAS significantly. Moreover, studies have reported that Gallic acid increases the activities of enzymes such as SOD, GPx and CAT, and increases GSH and vitamin C levels [33, 34]. Accordingly with the present study, it is thought that Gallic acid caused an increase in TAS levels by regulating nonenzymatic and/or enzymatic antioxidant mechanisms.

As another finding, when liver TOS values were compared, it decreased by 32 % in group I (13.3718 ± 1.13462) compared to the control group (19.6446 ± 2.41352) (P<0.5), whereas in group II (15.0483 ± 0.81577) a 23 % decrease was observed (P<0.5). However, it was determined that this value decreased by 14 % in group III (12.8825 ± 2.12539) compared to group II (P<0.5) (FIG. 2). In addition, liver OSI values decreased by 28 % in group I (13.7175) compared to the control group (19.0613), while it increased by 23 % in group II (23.5093). However, this value showed a 45 % regression in group III (12.9200) compared to group II (FIG. 3). The use of Gallic acid with H<sub>2</sub>O<sub>2</sub> has been shown to reduce liver TOS levels compared to hydrogen

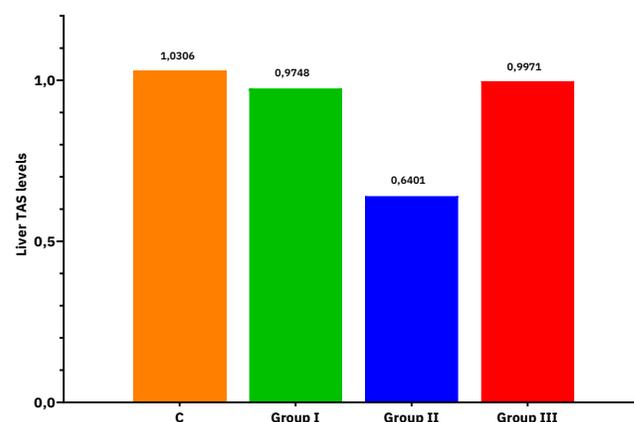
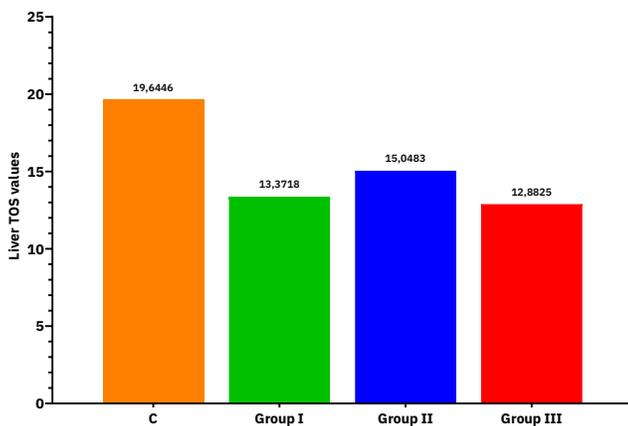


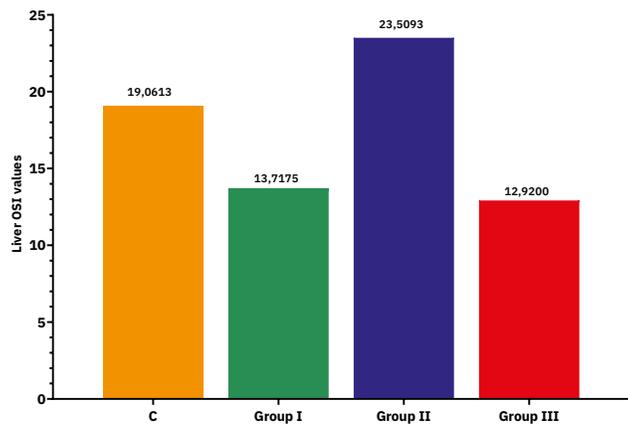
FIGURE 1. Total antioxidant status in liver homogenates of experimental groups (C: Control group, group I: only Gallic applied group, group II: only H<sub>2</sub>O<sub>2</sub> applied group, group III: gallic acid+H<sub>2</sub>O<sub>2</sub> applied group)

TABLE II  
Live weight values of experimental groups

	Initial live weight (g)	Final live weight (g)	10-day live weight change rate (%)	Average Daily Feed Intake Amount (g)
Control	210.52 ± 8.10	232.66 ± 6.93	10.52	33.40 ± 1.80
Group I (Gallic acid)	203.21 ± 5.88	212.26 ± 8.01	4.45	26.40 ± 4.80
Group II (H <sub>2</sub> O <sub>2</sub> )	210.82 ± 7.39	219.79 ± 9.24	4.25	36.40 ± 4.80
Group III (Gallic + H <sub>2</sub> O <sub>2</sub> )	210.16 ± 9.08	215.81 ± 10.92	2.69	24.40 ± 0.80
P	0.878	0.216		0.191



**FIGURE 2. Total oxidant status of experimental groups in liver homogenates (C: Control group, group I: only Gallic applied group, group II: only H<sub>2</sub>O<sub>2</sub> applied group, group III: gallic acid+H<sub>2</sub>O<sub>2</sub> applied group)**

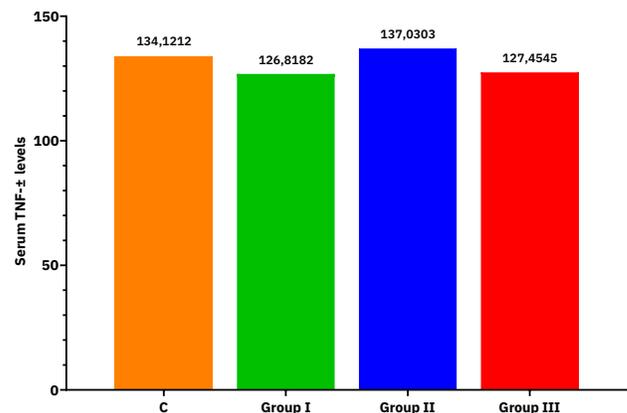


**FIGURE 3. Oxidative stress index values in liver homogenates of experimental groups (C: Control group, group I: only Gallic applied group, group II: only H<sub>2</sub>O<sub>2</sub> applied group, group III: Gallic acid+H<sub>2</sub>O<sub>2</sub> applied group)**

peroxide. Moreover, when the OSI values are considered, hydrogen peroxide increased the oxidative stress index, while the use of Gallic acid alone or in combination with H<sub>2</sub>O<sub>2</sub> significantly decreased the OSI levels. In a recent study, Ignea *et al.* [35] reported that Gallic acid GA increases the body's antioxidant capacity by directly neutralizing reactive oxygen species ROS and free radicals, or both, and reducing plasma malondialdehyde MDA levels. Jung *et al.* [36] reported that chicken breast meat was significantly larger in the group fed a mixture of 0.5% and 1.0% Gallic acid and Linoleic acid compared to the control group. They also reported that a mixture of Gallic acid and Linoleic acid were electron donors capable of neutralizing free radicals in broiler breast meat samples. Samuel *et al.* [32] reported that 100 mg/kg administration of Gallic acid significantly increased the broiler chest muscle mass compared to the control group, however, there was a decrease in the jejunal crypt depth and an increase in the villus width. Lee *et al.* [37] reported that the pH value of the broiler leg meat improved and the water holding capacity of the thigh increased in chickens fed with basal feed containing 1% Gallic acid for 2 weeks. In the same study, it was shown that Gallic acid increased the

antioxidant capacity of muscle tissues and significantly decreased the thiobarbituric acid reactive substances (TBARS) value. Jung *et al.* [38] showed that in chickens fed a mixture of Gallic acid and Linoleic acid, it reduced the cholesterol level of eggs, and also increased the antioxidant potential by reducing the TBARS level.

Inflammation is a natural, protective response of the organism to pathogens, chemical harmful stimuli or stress. As a physiological process, the main purpose of inflammatory responses is to induce the process of eliminating pathogens and toxins and repairing damaged tissue [39]. In the present study, blood TNF- $\alpha$  values decreased by 5% in group I (126.8182 $\pm$ 21.06224) compared to the control group (134.1212 $\pm$ 21.14828) ( $P>0.5$ ), while group II (137.0303 $\pm$ 23.78826) showed an increase of 2% ( $P<0.5$ ). However, this value showed a 7% regression in group III (127.4545 $\pm$ 12.26777) compared to group II ( $P<0.5$ ) (FIG. 4). Oxidative stress is closely associated with inflammation due to the fact that the NF- $\kappa$ B pathway, which is important in the regulation of inflammation, is activated due to the increase of reactive oxygen species [40]. It has been reported that increased ROS may cause nuclear factor kappa B (NF- $\kappa$ B) activation and expression of inflammatory cytokines [41]. Therefore, according to the results obtained, it is thought that Gallic acid may have shown this effect by reducing ROS with its strong antioxidant property.



**FIGURE 4. Effects of Gallic acid and Hydrogen peroxide on TNF- $\alpha$  translation levels in serum samples (C: Control group, group I: only Gallic applied group, group II: only H<sub>2</sub>O<sub>2</sub> applied group, group III: Gallic acid+H<sub>2</sub>O<sub>2</sub> applied group)**

It was found that while H<sub>2</sub>O<sub>2</sub> induced blood TNF- $\alpha$  gene expression levels of Japanese quails, Gallic acid significantly suppressed the expression levels of this gene. It was seen that there are studies investigating the anti-inflammatory potential of Gallic acid. In a study investigating the effects of polyphenols in broilers, it was determined that tea polyphenols (0.03-0.09 g·kg<sup>-1</sup> body weight) in the long term were found to show anti-inflammatory activity by reducing gene expression levels of proinflammatory cytokines such as IL-1 $\beta$ , IL-4, IL-6, IL-10, TNF- $\alpha$  and IFN- $\gamma$  [39].

## CONCLUSIONS

In the present study, Gallic acid application did not show a statistically significant change on live weight change rates of

Japanese quails. However, it increased liver TAS levels suppressed by hydrogen peroxide and decreased liver TOS levels in quails. Considering the total OSI values, it can be said that it makes a significant contribution to suppressing oxidative stress. In addition, it showed anti-inflammatory activity by significantly suppressing TNF- $\alpha$ , one of the proinflammatory cytokines. As a result, in terms of its antioxidant and anti-inflammatory activities, Gallic acid is a compound that may have antioxidant and anti-inflammatory potentials in Japanese quails. However, further and detailed studies are needed to fully elucidate this argument.

### Conflict of interests

The authors of this study declare that there is no conflict of interest with the publication of this manuscript.

### BIBLIOGRAPHICS REFERENCES

- [1] Gadde U, Kim WH, Oh ST, Lillehoj HS. Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry: a review. *Anim. Health Res. Rev.* 2017; 18(1):26–45.
- [2] Lee MT, Lin WC, Lee TT. Potential crosstalk of oxidative stress and immune response in poultry through phytochemicals – A review. *Asian–Australasian J. Anim. Sci.* 2019; 32(3):309–19.
- [3] Kumar N, Goel N. Phenolic acids: Natural versatile molecules with promising therapeutic applications. *Biotechnol. Rep. (Amst).* 2019; 24:e00370.
- [4] Siah M, Farzaei MH, Ashrafi–Kooshk MR, Adibi H, Arab SS, Rashidi MR, Khodarahmi R. Inhibition of guinea pig aldehyde oxidase activity by different flavonoid compounds: An *in vitro* study. *Bioorganic. Chem.* 2016; 64:74–84.
- [5] Kahkeshani N, Farzaei F, Fotouhi M, Alavi SS, Bahramsoltani R, Naseri R, Momtaz S, Abbasabadi Z, Rahimi R, Farzaei MH, Bishayee A. Pharmacological effects of gallic acid in health and disease: A mechanistic review. *Iranian J. Basic Med. Sci.* 2019; 22(3):225–237. doi: <https://doi.org/gng34z>
- [6] Fernandes FHA, Salgado HRN. Gallic acid: review of the methods of determination and quantification. *Critic. Rev. Analytical Chem.* 2016; 46(3):257–65.
- [7] Hsieh HM, Ju YM. Medicinal components in *Termitomyces mushrooms*. *Appl. Microbiol. Biotechnol.* 2018; 102(12):4987–94.
- [8] Zhang T, Ma L, Wu P, Li W, Li T, Gu R, Dan X, Li Z, Fan X, Xiao Z. Gallic acid has anticancer activity and enhances the anticancer effects of cisplatin in non-small cell lung cancer A549 cells via the JAK/STAT3 signaling pathway. *Oncol. Rep.* 2019; 41(3):1779–1788.
- [9] Rivero–Buceta E, Carrero P, Doyagüez EG, Madrona A, Quesada E, Camarasa MJ, Pérez–Pérez MJ, Leyssen P, Paeshuyse J, Balzarini J, Neyts J, San–Félix A. Linear and branched alkyl–esters and amides of gallic acid and other (mono-, di- and tri-) hydroxy benzoyl derivatives as promising anti–HCV inhibitors. *Europ. J. Med. Chem.* 2015; 92:656–71.
- [10] Jung J, Bae KH, Jeong CS. Anti–*Helicobacter pylori* and antiulcerogenic activities of the root cortex of *Paeonia suffruticosa*. *Biol. Pharmac. Bull.* 2013; 36(10):1535–9.
- [11] Couto AG, Kassuya CAL, Calixto JB, Petrovick PR. Anti-inflammatory, antiallodynic effects and quantitative analysis of gallic acid in spray dried powders from *Phyllanthus niruri* leaves, stems, roots and whole plant. *Rev. Brasileira Farmacognosia.* 2013; 23(1):124–31.
- [12] Sarjit A, Wang Y, Dykes GA. Antimicrobial activity of gallic acid against thermophilic *Campylobacter* is strain specific and associated with a loss of calcium ions. *Food Microbiol.* 2015; 46:227–33.
- [13] Li ZJ, Liu M, Dawuti G, Dou Q, Ma Y, Liu HG, Aibai S. Antifungal activity of gallic acid *in vitro* and *in vivo*. *Phyther. Res.* 2017; 31(7):1039–45.
- [14] Badavi M, Sadeghi N, Dianat M, Samarbafzadeh A. Effects of gallic acid and cyclosporine a on antioxidant capacity and cardiac markers of rat isolated heart after ischemia/reperfusion. *Iranian Red Crescent Med. J.* 2014; 16(6):1–7.
- [15] Estévez M. Oxidative damage to poultry: from farm to fork. *Poult. Sci.* 2015; 94(6):1368–78.
- [16] Zhang YJ, Gan RY, Li S, Zhou Y, Li AN, Xu DP, Li HB. Antioxidant phytochemicals for the prevention and treatment of chronic diseases. *Molecules.* 2015; 20(12):21138–56.
- [17] Hu R, He Y, Arowolo M, Wu S, He J. Polyphenols as potential attenuators of heat stress in poultry production. *Antioxid.* 2019; 8(3):67.
- [18] Kucukgul A, Erdogan S. Caffeic acid phenethyl ester (CAPE) protects lung epithelial cells against H<sub>2</sub>O<sub>2</sub>–induced inflammation and oxidative stress. *Health Med.* 2014; 8(3):329–338.
- [19] Abdel–Wahab A, Abdel–Kader I, Ahmad E. Effect of dietary grape seed supplementation as a natural growth promoter on the growth performance of japanese quail. *Egypt. J. Nutr. Feeds.* 2018; 21(2):537–48.
- [20] Silici S, Güçlü BK, Kara K. Yumurtacı damızlık bıldırcın (*Coturnix coturnix japonica*) yemlerine öğütülmüş üzüm çekirdeği ilavesinin verim ve kuluçka performansı ile yumurta kalitesine etkisi. *ERU Sağlık Bilimleri Dergisi.* 2011; 20(1):68–76.
- [21] Abu Hafsa SH, Ibrahim SA. Effect of dietary polyphenol-rich grape seed on growth performance, antioxidant capacity and ileal microflora in broiler chicks. *J. Anim. Physiol. Anim. Nutr.* 2018; 102(1):268–75.
- [22] Ao X, Kim IH. Effects of grape seed extract on performance, immunity, antioxidant capacity, and meat quality in Pekin ducks. *Poult. Sci.* 2020; 99(4):2078–86.
- [23] Surai PF, Kochish II, Fisinin VI, Kidd MT. Antioxidant defence systems and oxidative stress in poultry biology: an update. *Antioxid.* 2019; 8(7):235.
- [24] Lee MT, Lin WC, Yu B, Lee TT. Antioxidant capacity of phytochemicals and their potential effects on oxidative status in animals – A review. *Asian–Australasian J. Anim. Sci.* 2017; 30(3):299–308.
- [25] Surai PF, Kochish II. Nutritional modulation of the antioxidant capacities in poultry: the case of selenium. *Poult. Sci.* 2019; 98(10):4231–9.
- [26] Saibabu V, Fatima Z, Khan LA, Hameed S. Therapeutic potential of dietary phenolic acids. *Adv. Pharmacol. Sci.* 2015; 2015:823539.

- [27] Makihara H, Koike Y, Ohta M, Horiguchi-Babamoto E, Tsubata M, Kinoshita K, Akase T, Goshima Y, Aburada M, Shimada T. Gallic acid, the active ingredient of *Terminalia bellirica*, enhances adipocyte differentiation and adiponectin secretion. *Biol. Pharmac. Bull.* 2016; 39(7):1137-43.
- [28] Gandhi GR, Jothi G, Antony PJ, Balakrishna K, Paulraj MG, Ignacimuthu S, Stalin A, Al-Dhabi NA. Gallic acid attenuates high-fat diet fed-streptozotocin-induced insulin resistance via partial agonism of PPAR $\gamma$  in experimental type 2 diabetic rats and enhances glucose uptake through translocation and activation of GLUT4 in PI3K/p-Akt signaling pathway. *Europ. J. Pharmacol.* 2014; 745:201-16.
- [29] Totani N, Tateishi S, Takimoto T, Maeda Y, Sasaki H. Gallic acid glycerol ester promotes weight-loss in rats. *J. Oleo Sci.* 2011; 60(9):457-62.
- [30] Locatelli C, Filippin-Monteiro FB, Creczynski-Pasa TB. Alkyl esters of gallic acid as anticancer agents: A review. *Europ. J. Med. Chem.* 2013; 60:233-9.
- [31] Choubey S, Varughese LR, Kumar V, Beniwal V. Medicinal importance of gallic acid and its ester derivatives: a patent review. *Pharmac. Patent Analyst.* 2015; 4(4):305-15.
- [32] Samuel KG, Wang J, Yue HY, Wu SG, Zhang HJ, Duan ZY, Qi GH. Effects of dietary gallic acid supplementation on performance, antioxidant status, and jejunum intestinal morphology in broiler chicks. *Poult. Sci.* 2017; 96(8):2768-75.
- [33] Nouri A, Heibati F, Heidarian E. Gallic acid exerts anti-inflammatory, anti-oxidative stress, and nephroprotective effects against paraquat-induced renal injury in male rats. *Naunyn Schmiedebergs Arch Pharmacol.* 2021; 394(1):1-9.
- [34] Ahmadvand H, Yalameha B, Adibhesami G, Nasri M, Naderi N, Babaeenezhad E, Nouryazdan N. The Protective Role of Gallic Acid Pretreatment On Renal Ischemia-reperfusion Injury in Rats. *Rep. Biochem. Mol. Biol.* 2019;8(1):42-48.
- [35] Ignea C, Dorobanțu CM, Mintoff CP, Branza-Nichita N, Ladomery MR, Kefalas P, Chedea VS. Modulation of the antioxidant/pro-oxidant balance, cytotoxicity and antiviral actions of grape seed extracts. *Food Chem.* 2013; 141(4):3967-76
- [36] Jung S, Choe JH, Kim B, Yun H, Kruk ZA, Jo C. Effect of dietary mixture of gallic acid and linoleic acid on antioxidative potential and quality of breast meat from broilers. *Meat Sci.* 2010; 86(2):520-6.
- [37] Lee KH, Jung S, Kim HJ, Kim IS, Lee JH, Jo C. Effect of dietary supplementation of the combination of gallic and linoleic acid in thigh meat of broilers. *Asian-Australasian J. Anim. Sci.* 2012; 25(11):1641-8.
- [38] Jung S, Han BH, Nam K, Ahn DU, Lee JH, Jo C. Effect of dietary supplementation of gallic acid and linoleic acid mixture or their synthetic salt on egg quality. *Food Chem.* 2011; 129(3):822-9.
- [39] Medzhitov R. Inflammation 2010: New adventures of an old flame. *Cell.* 2010; 140(6):771-6.
- [40] Pantano C, Reynaert NL, Vliet AVD, Janssen-Heininger YMW. Redox-sensitive kinases of the nuclear factor- $\kappa$ B signaling pathway. *Antioxid. Redox Signaling.* 2006; 8(9-10):1791-806.
- [41] Li HL, Li ZJ, Wei ZS, Liu T, Zou XZ, Liao Y, Luo Y. Long-term effects of oral tea polyphenols and *Lactobacillus brevis* M8 on biochemical parameters, digestive enzymes, and cytokines expression in broilers. *J. Zhejiang University-Science B.* 2015; 16(12):1019-26.